# Unexpected infection spikes in a model of Respiratory Syncytial Virus vaccination

Robert J. Smith?<sup>1</sup>, Alexandra Hogan<sup>2</sup>, Geoff Mercer<sup>2</sup>

1. Department of Mathematics, The University of Ottawa, 585 King Edward Ave, Ottawa ON K1N 6N5 Canada

2. National Centre for Epidemiology and Population Health, Australian National University, Canberra, 2601, Australia

#### Abstract

Respiratory Syncytial Virus (RSV) is an actute respiratory infection that infects millions of children and infants worldwide. Recent research has shown promise for the development of live attentuated vaccines, several of which are in clinical trials or preclinical development. We extend an existing mathematical model with seasonal transmission to include vaccination. We model vaccination both as a continuous process and as a discrete one, using impulsive differential equations. We develop conditions for the stability of the disease-free equilibrium and show that this equilibrium can be destabilised under certain (extreme) conditions. Using impulsive differential equations and introducing a new quantity, the *impulsive reproduction number*, we determine conditions for the period and strength of vaccination that will control (but not eradicate) RSV. The waning rate of the vaccine is a critical parameter for long-term reduction in RSV prevalence, even more than coverage. We recommend that candidate vaccines be tested for sufficient duration before being released on the market.

## 1 Introduction

Respiratory syncytial virus (RSV) is the main cause of acute lower respiratory infections in infants and young children [27], with almost all children having been infected by two years of age [14, 31] and an estimated 0.5–2% of infants requiring hospitalisation due to infection [23]. One recent study estimated that, in 2005, 33.8 million new episodes of RSV occurred worldwide in children younger than five years of age [27]. Symptoms of RSV range from those of a cold to more severe afflictions such as bronchiolitis and pneumonia [14]. While mortality due to RSV infection in developed countries is low, occurring in less than 0.1% of cases [38], few data have been published about RSV morbidity and mortality in developing countries [40]. However, estimates of the hospitalisation costs are substantial [18, 36, 42], making RSV a significant economic and health care system burden.

Newborn infants are typically protected from RSV infection by maternal antibodies until about six weeks of age [12], and the highest number of observed RSV cases occur in children aged six weeks to six months [8, 33]. Immunity to RSV following an infection is short-lasting, and reinfection in childhood is common [24]. Few studies have been undertaken to investigate transmission of RSV among adults, but it is thought that infection can occur throughout life [9, 19] and that, in older children and adults, RSV manifests as a mild cold [14, 20]. RSV has been identified as a cause of mortality in the elderly, with documented outbreaks in aged-care settings [17, 37]; one such study found that up to 18% of pneumonia hospitalisation in adults aged above 65 years may be due to RSV infection [16].

In temperate climates RSV epidemics exhibit distinct and consistent seasonal patterns. Most RSV infections occur during the cooler winter months, whether wet or dry [40], and outbreaks typically last between two and five months [15, 28]. In a number of temperate regions, a biennial pattern for RSV cases has been identified [4, 25, 34]. In tropical climates, RSV is detected throughout the year with less pronounced seasonal peaks, and the onset of RSV is typically associated with the wet season [32, 40].

Immunoprophylaxis with the monoclonal antibody Palivizumab, while not preventing the onset of infection, has proven effective in reducing the severity of RSV-related symptoms [35]. However, prophylaxis is expensive and generally only administered to high-risk children, with recommendations varying across jurisdictions. There is currently no licensed vaccine to prevent RSV infection, despite about 50 years of vaccine research. Recent research has focused on the development of live attenuated vaccines; several such vaccines are being evaluated in clinical trials, with other vaccines in preclinical development [13, 18]. With the possibility of an RSV vaccine becoming available, mathematical models can be powerful tools for planning vaccination roll-out strategies.

Several ordinary differential equation mathematical models for RSV transmission have been published to date [3, 10, 22, 26, 29, 39, 41], most using Susceptible–Exposed–Infectious–Recovered (SEIR) dynamics and with a sine or cosine forcing term to account for seasonal variation in transmission. Weber *et al.* [39] presented an SEIRS model that incorporated a gradual reduction in susceptibility to reinfection and maternally derived immunity; they fitted the model to several data sets. Leecaster *et al.* [22] presented an SEIDR model with both child and adult classes for the S, E and I compartments, where the D class represented children in which infection was detected. The model was fitted to seven years of data from Salt Lake City, USA.

Moore *et al.* [26] presented an age-structured SEIRS model for children under two years of age, as well as the remaining population. The model was fitted to data from Perth, Western Australia. Capistran *et al.* [10] outlined an SIRS model with seasonal forcing and proposed a method to estimate the model parameters, demonstrated by fitting models to data from The Gambia and Finland. Paynter *et al.* [29] investigated the ecological drivers of RSV seasonality in the Philippines, where the model included a second partially susceptible class and classes for latent and infectious individuals with subsequent RSV infection. They applied a square-wave transmission term that accounted for decreased transmissibility over the summer holidays, as well as a seasonally driven birth rate.

White *et al.* [41] used nested differential equation models to describe RSV transmission and fitted these to RSV case data for eight different regions. Arenas *et al.* [3] introduced randomness into the differential equation model and fitted the model to RSV hospitalisation data for Valencia, Spain.

Few papers have so far explored vaccination strategies for RSV. A newborn vaccination strategy was outlined in Acedo *et al.* [1] for the Spanish region of Valencia, in order to estimate the cost-effectiveness of potential RSV vaccination strategies. Their modelling approach removed a fraction of susceptible newborns into a vaccinated class, where they remained until they reached the next age group, at which point they moved to the second susceptible class. This strategy assumes booster doses of the vaccine in the first year of life, such that the immunisation period would be at least equal to the immunity of those who have recovered from RSV infection. In subsequent work, an RSV vaccine cost analysis was conducted based on a stochastic network model, with children vaccinated at two months, four months and one year of age [2].

Here, we examine the effects of a theoretical vaccine on the spread of RSV. We examine several vaccination scenarios, including differing levels of coverage, seasonal oscillations in the transmission rate and a waning of the vaccine. We also compare continuous vaccination to impulsive vaccination in order to determine conditions on the vaccination strength and period that will control the virus.

# 2 The model

We extend the basic model from Weber *et al.* [39] to include vaccination. We assume that the leaving rate  $\mu$  is unchanged across all classes and that there is no disease-specific death rate. We scale the entry and leaving rates so that the population is constant.

Let S represent susceptible, I represent infected and R represent recovered individuals, with V,  $I_V$  and  $R_V$  the corresponding compartments for vaccinated individuals. The birth rate is  $\mu$ , with a proportion p vaccinated, of whom  $\epsilon$  successfully mount an immune response; the death rate is matched to the birth rate. The time-dependent transmissibility parameter is  $\beta(t)$ , with recovery  $\nu$  and loss of immunity  $\gamma$ . Corresponding vaccination parameters are  $\beta_V(t)$ ,  $\nu_V$  and  $\gamma_V$ , respectively. Finally, the waning of the vaccine is given by  $\omega$ .

The basic model with vaccination is then

$$S' = \mu(1 - \epsilon p) - \mu S - \beta(t)S(I + I_V) + \gamma R + \omega V$$
$$I' = \beta(t)S(I + I_V) - \nu I - \mu I + \omega I_V$$
$$R' = \nu I - \mu R - \gamma R + \omega R_V$$
$$V' = \epsilon p \mu - \mu V - \beta_V(t)V(I + I_V) + \gamma_V R_V - \omega V$$
$$I'_V = \beta_V(t)V(I + I_V) - \nu_V I_V - \mu I_V - \omega I_V$$
$$R'_V = \nu_V I_V - \mu R_V - \gamma_V R_V - \omega R_V,$$

with  $\beta(t) = b_0(1 + b_1 \cos(2\pi t + \phi))$  and  $\beta_V(t) = (1 - \alpha)\beta(t)$ , for  $0 \le \alpha \le 1$ . (We may relax the lower bound on  $\alpha$  later.) The model is illustrated in Figure 1.

# 3 Analysis

There is a disease-free equilibrium that satisfies

$$(\bar{S}, \bar{I}, \bar{R}, \bar{V}, \bar{I}_V, \bar{R}_V) = \left(\frac{(1-\epsilon p)\mu + \omega}{\mu + \omega}, 0, 0, \frac{\epsilon p\mu}{\mu + \omega}, 0, 0\right).$$



Figure 1: The model.

## 3.1 Constant transmission

If we assume transmission is constant, so that  $\beta$  and  $\beta_V$  are independent of time, then the Jacobian is  $J = [J_1|J_2]$ , where

$$J_{1} = \begin{bmatrix} -\mu - \beta(I + I_{V}) & -\beta \bar{S} & \gamma \\ \beta(I + I_{V}) & \beta \bar{S} - \mu - \nu & 0 \\ 0 & \nu & -\mu - \gamma \\ 0 & -\beta_{V} \bar{V} & 0 \\ 0 & 0 & 0 \end{bmatrix}$$
$$J_{2} = \begin{bmatrix} \omega & -\beta \bar{S} & 0 \\ 0 & \beta \bar{S} + \omega & 0 \\ 0 & 0 & \omega \\ -\mu - \beta_{V} (I + I_{V}) - \omega & -\beta_{V} \bar{V} & \gamma_{V} \\ \beta_{V} (I + I_{V}) & \beta_{V} \bar{V} - \nu_{V} - \mu - \omega & 0 \\ 0 & \nu_{V} & -\mu - \gamma_{V} - \omega \end{bmatrix}.$$

At the DFE, we have

$$J\Big|_{\mathrm{D}FE} = \begin{bmatrix} -\mu & -\beta \bar{S} & \gamma & \omega & -\beta \bar{S} & 0\\ 0 & \beta \bar{S} - \mu - \nu & 0 & 0 & \beta \bar{S} + \omega & 0\\ 0 & \nu & -\mu - \gamma & 0 & 0 & \omega\\ 0 & -\beta_V \bar{V} & 0 & -\mu - \omega & -\beta_V \bar{V} & \gamma_V\\ 0 & \beta_V \bar{V} & 0 & 0 & \beta_V \bar{V} - \nu_V - \mu - \omega & 0\\ 0 & 0 & 0 & 0 & \nu_V & -\mu - \gamma_V - \omega \end{bmatrix}.$$

The characteristic polynomial satisfies

$$\det(J - \lambda I) = (-\mu - \lambda)(-\mu - \gamma - \lambda)(-\mu - \omega - \lambda)(-\mu - \gamma_V - \omega - \lambda) \det M,$$
  
where

$$M = \begin{bmatrix} \beta \bar{S} - \mu - \nu - \lambda & \beta \bar{S} + \omega \\ \beta_V \bar{V} & \beta_V \bar{V} - \nu_V - \mu - \omega - \lambda \end{bmatrix}$$

The first four eigenvalues are always negative. The nontrivial part of characteristic equation satisfies

$$\lambda^2 + b_1 \lambda + c_1 = 0,$$

where

$$b_1 = -\beta \bar{S} + \mu + \nu - \beta_V \bar{V} + \nu_V + \mu + \omega$$
  

$$c_1 = (\beta \bar{S} - \mu - \nu)(\beta_V \bar{V} - \nu_V - \mu - \omega) - \beta_V \bar{V}(\beta \bar{S} + \omega)$$
  

$$= \beta \bar{S}(-\nu_V - \mu - \omega) - (\mu + \nu)(\beta_V \bar{V} - \nu_V - \mu - \omega) - \beta_V \bar{V}\omega.$$

From  $c_1 = 0$ , we find

$$R_0 = \frac{\beta \bar{S}(\nu_V + \mu + \omega) + \beta_V \bar{V}(\mu + \nu + \omega)}{(\mu + \nu)(\mu + \nu_V + \omega)}$$

(This is equivalent to the value found using the next-generation method.)

If  $c_1 = 0$  and  $b_1 > 0$ , then we have a bifurcation with the property that the DFE is stable if  $R_0 < 1$  and unstable if  $R_0 > 1$ 

However, it is possible that when  $c_1 = 0$ ,  $b_1 < 0$ . In this case,  $R_0$  is not a threshold and the disease can persist if  $R_0 < 1$ .

When  $c_1 = 0$ , we have

$$b_1 \Big|_{c_1=0} = \frac{1}{\nu_V + \mu + \omega} \left[ \beta_V \bar{V}(\nu - \nu_V) + (\nu_V + \mu + \omega)^2 \right].$$

Note that if  $\nu = \nu_V$ , then  $b_1 > 0$ . However, we expect that vaccinated individuals will recover faster than unvaccinated individuals. Thus  $\nu_V > \nu$ . This raises the possibility that  $b_1$  could be negative.

If  $\nu_V \to \infty$ , then this is equivalent to vaccinated individuals recovering instantaneously. In this case,

$$\lim_{\nu_V \to \infty} b_1 = \lim_{\nu_V \to \infty} \frac{\beta_V V(\nu - \nu_V)}{\omega + \mu + \nu_V} + \omega + \mu + \nu_V$$
$$= \infty - \beta_V \overline{V} > 0$$



Figure 2: Possible sketch of the form of  $f(\nu_V)$  with a negative minimum between two positive extremes.

Hence if we define  $f(\nu_V) = \frac{\beta_V \bar{V}(\nu - \nu_V) + (\omega + \mu + \nu_V)^2}{\omega + \mu + \nu_V}$ , then it is clear that f(0) > 0 and  $f(\infty) > 0$ . So we would like to know whether f has a turning point  $\nu_V^*$  such that  $f(\nu_V^*) < 0$ .

We have

$$f'(\nu_V) = \frac{(\omega + \mu + \nu_V)[-\beta_V V + 2(\omega + \mu + \nu_V)] - [\beta_V V(\nu - nu_V) + (\omega + \mu + \nu_V)^2]}{(\omega + \mu + \nu_V)^2}$$
$$= \frac{(\omega + \mu + \nu_V)^2 - \beta_V \bar{V}[\omega + \mu + \nu_V)^2}{(\omega + \mu + \nu_V)^2}$$

It follows that  $\nu_V^* = \sqrt{\beta_V \bar{V}(\omega + \mu + \nu)} - \omega - \mu$ . There are three requirements we need for this to be meaningful:

- 1.  $\nu_V^* > \nu$
- 2.  $f(\nu_V^*) < 0$  and
- 3.  $\nu_V^*$  is a local minimum.

See Figure 2.

The first and second criteria determine whether such a  $\nu_V^*$  exists. To prove the third, we can differentiate again:

$$f''(\nu_V) = \frac{(\omega + \mu + \nu_V)^2 + \beta_V(\omega + \mu + \nu)}{(\omega + \mu + \nu_V)} > 0.$$

It follows that  $\nu_V^*$  is a local minimum whenever it exists.

# 4 Impulsive model

Previously, we assumed that vaccination occurred at birth and that a fixed proportion of newborns were vaccinated. This is effectively continuous vaccination. However, vaccination may occur later and may be administered at regular times (for example, in schools or daycare centres). We assume that the effect of the vaccine is to reduce the susceptible population by a fixed proportion. Such a model is described by a system of non-autonomous impulsive differential equations [5, 6, 7, 21].

The impulsive model is given by

$S' = \mu - \mu S - \beta(t)S(I + I_V) + \gamma R + \omega V$	$t \neq t_k$
$I' = \beta(t)S(I+I_V) - \nu I - \mu I + \omega I_V$	$t \neq t_k$
$R' = \nu I - \mu R - \gamma R + \omega R_V$	$t \neq t_k$
$V' = -\mu V - \beta_V(t)V(I + I_V) + \gamma_V R_V - \omega V$	$t \neq t_k$
$I'_V = \beta_V V(I + I_V) - \nu_V I_V - \mu I_V - \omega I_V$	$t \neq t_k$
$R'_V = \nu_V I_V - \mu R_V - \gamma_V R_V - \omega R_V$	$t \neq t_k$
$\Delta S = -rS$	$t = t_k$
$\Delta V = rS$	$t = t_k$

Here  $t_k$  are the vaccination times. They may be fixed or non-fixed, although for our purposes we will consider them fixed.

#### 4.1 Impulsive Analysis

We will set  $\beta$  to be constant for mathematical convenience. In order to analyse the impulsive system, we need to solve the differential equations for finite time. Since this is not possible in general, we will develop several overestimates in order to determine bounds for the long-term numbers of susceptible, infected and vaccinated individuals, under a few assumptions.

#### Susceptible individuals

First we consider the overestimate  $I + I_V \leq 1$  (i.e., maximal infection). Then we have

$$S' \ge \mu - \mu S - \beta S.$$

Integrating and applying the "initial" condition  $S(t_k^+)$  in the (k+1)st cycle,

we have

$$S(t) \ge e^{-(\mu+\beta)(t-t_k)}S(t_k^+) + \frac{\mu}{\mu+\beta} \left(1 - e^{-(\mu+\beta)(t-t_k)}\right), \text{ for } t_k < t \le t_{k+1}$$
$$S(t_{k+1}^-) \ge e^{-(\mu+\beta)\tau}S(t_k^+) + \frac{\mu}{\mu+\beta} \left(1 - e^{-(\mu+\beta)\tau}\right).$$

Applying the impulsive condition, we have

$$S(t_{k+1}^+) = (1-r)S(t_k^-)$$
  

$$S(t_{k+1}^+) \ge (1-r)e^{-(\mu+\beta)\tau}S(t_k^+) + \frac{\mu}{\mu+\beta}(1-r)\left(1-e^{-(\mu+\beta)\tau}\right).$$

This is a recurrence relation in the form  $x_{n+1} = ax_n + b$ , which has equilibrium  $\bar{x} = \frac{b}{1-a}$ , and the equilibrium is stable if |a| < 1. In our case, we have  $a = (1-r)e^{-(\mu+\beta)\tau} < 1$ , so the equilibrium is stable. It follows that solutions converge to a stable impulsive periodic orbit with endpoints

$$S_{\infty}^{-} = \frac{\mu \left(1 - e^{-(\mu+\beta)\tau}\right)}{(\mu+\beta) \left(1 - (1-r)e^{-(\mu+\beta)\tau}\right)}$$
$$S_{\infty}^{+} = \frac{\mu (1-r) \left(1 - e^{-(\mu+\beta)\tau}\right)}{(\mu+\beta) \left(1 - (1-r)e^{-(\mu+\beta)\tau}\right)}.$$

These values correspond to the local maximum and minimum values for the unvaccinated susceptibles after a long time. These values are well-defined, since both the numerator and the denominator are always positive.

Note in particular that

$$\lim_{\tau \to 0} S_{\infty}^{-} = 0.$$

That is, if the period between vaccinations shrinks to zero, then the number of susceptibles would shrink to zero. (Note that this is a theoretical result only, since the impulsive assumptions of long cycle times relative to instantaneous approximation would break down [11].)

#### Vaccinated individuals

Second, we turn our attention to vaccination. Using the inequalities  $I + I_V \leq 1$  and  $R_V \geq 0$ , we have

$$V' \ge -\mu V - \beta V - \omega V.$$

Integrating and applying the "initial" condition  $V(t_k^+)$  in the  $(k+1) \mathrm{st}$  cycle, we have

$$V(t) \ge V(t_k^+) e^{-(\mu + \beta + \omega)(t - t_k)}, \text{ for } t_k < t \le t_{k+1}$$
$$V(t_{k+1}^-) \ge V(t_k^+) e^{-(\mu + \beta + \omega)\tau}.$$

Applying the impulsive condition, we have

$$\begin{split} V(t_{k+1}^+) &= V(t_{k+1}^-) + rS(t_{k+1}^1) \\ V(t_{k+1}^+) &\geq V(t_{k+1}^-) + \frac{r\mu \left(1 - e^{-(\mu+\beta)\tau}\right)}{(\mu+\beta) \left(1 - (1-r)e^{-(\mu+\beta)\tau}\right)} \\ &\geq V(t_k^-)e^{-(\mu+\beta+\omega)\tau} + \frac{r\mu \left(1 - e^{-(\mu+\beta)\tau}\right)}{(\mu+\beta) \left(1 - (1-r)e^{-(\mu+\beta)\tau}\right)}. \end{split}$$

Since  $e^{-(\mu+\beta+\omega)\tau} < 1$ , this recurrence relation has a stable equilibrium and hence solutions converge to the impulsive periodic orbit with endpoints

$$V_{\infty}^{-} = \frac{r\mu \left(1 - e^{-(\mu+\beta)\tau}\right) e^{-(\mu+\beta+\omega)\tau}}{(\mu+\beta) \left(1 - (1-r)e^{-(\mu+\beta)\tau}\right) \left(1 - e^{(\mu+\beta+\omega)\tau}\right)}$$
$$V_{\infty}^{+} = \frac{r\mu \left(1 - e^{-(\mu+\beta)\tau}\right)}{(\mu+\beta) \left(1 - (1-r)e^{-(\mu+\beta)\tau}\right) \left(1 - e^{(\mu+\beta+\omega)\tau}\right)}.$$

#### Infected vaccinated individuals

Next we calculate the number of infected vaccinated individuals. Using the overestimate  $I \leq 1,$  we can write

$$I' \leq \beta_V V(1+I_V) - \nu_V I_V - \mu I_V - \omega I_V$$
  
$$\leq \beta_V V_{\infty}^+(1+I_V) - \nu_V I_V - \mu I_V - \omega I_V$$

in the long run. Integrating and applying the initial condition I(0) = 0, we have

$$I_V = \frac{\beta V_\infty^+}{\nu_V + \mu + \omega - \beta_V V_\infty^+} \left( 1 - e^{(\beta_V V_\infty^+ - \nu_V - \mu - \omega)t} \right).$$

This converges if  $\nu_V + \mu + \omega - \beta_V V_{\infty}^+ > 0$ . If this holds, then

$$\nu_V + \mu + \omega > \frac{\beta_V r \mu \left(1 - e^{-(\mu + \beta)\tau}\right)}{(\mu + \beta) \left(1 - (1 - r)e^{-(\mu + \beta)\tau}\right) \left(1 - e^{(\mu + \beta + \omega)\tau}\right)}.$$

Rearranging, we have

$$(\mu + \beta) \left( 1 - (1 - r)e^{-(\mu + \beta)\tau} \right) (\nu_V + \mu + \omega) > \beta_V r \mu \left( 1 - e^{-(\mu + \beta)\tau} \right)$$
$$[-(\mu + \beta)(1 - r)(\nu_V + \mu + \omega) + \beta_V r \mu] e^{-(\mu + \beta)\tau} > \beta_V r \mu - (\mu + \beta)(\nu_V + \mu + \omega).$$

This inequality has no solution unless

$$r < \bar{r} = \frac{(\mu + \beta)(\nu_V + \mu + \omega)}{\beta_V r \mu + (\mu + \beta)(\nu_V + \mu + \omega)} < 1$$

If  $r > \bar{r}$ , then  $I_V$  converges to

$$\lim_{t \to \infty} I_V = \frac{\beta_V V_\infty^+}{\nu_V + \mu + \omega - \beta_V V_\infty^+} \equiv I_V^\infty$$

We are interested in the size of this value. We have

$$I_V^{\infty} = \frac{\beta_V r \mu \left(1 - e^{-(\mu + \beta)\tau}\right)}{\mu + \beta \left(1 - (1 - r)e^{-(\mu + \beta)\tau}\right) \left(1 - e^{-(\mu + \beta + \omega)\tau}\right) \left(\nu_V + \mu + \omega - \frac{\beta_V r \mu \left(1 - e^{-(\mu + \beta)\tau}\right)}{(\mu + \beta) \left(1 - (1 - r)e^{-(\mu + \beta)\tau}\right)}\right)}$$

To estimate the size of this value for frequent vaccinations, we use L'Hôpital's rule to find

$$\lim_{\tau \to 0} I_V^{\infty} = \frac{\beta_V \mu}{(\mu + \beta + \omega)(\nu_V + \mu + \omega)}$$
$$= \frac{(1 - \alpha)\beta\mu}{(\mu + \beta + \omega)(\nu_V + \mu + \omega)} \ll 1 - \alpha < 1.$$

It follows that  $I_V$  is small if the vaccine significantly reduces transmissibility and is applied frequently.

#### Infected individuals

Finally, we examine the number of infected individuals under the assumption that infected vaccinated individuals are negligible (so  $I_V \approx 0$ ). We then have

$$\begin{split} I' &\approx \beta SI - \nu I - \mu I \\ &\leq \beta S_{\infty}^{-}I - \nu I - \mu I \\ &= \frac{\beta \mu \left(1 - e^{-(\mu + \beta)\tau}\right)}{(\mu + \beta) \left(1 - (1 - r)e^{-(\mu + \beta)\tau}\right)} I - \nu I - \mu I. \end{split}$$

It follows that, after sufficient time, the disease will be contracting if

$$q = \frac{\beta \mu \left( 1 - e^{-(\mu + \beta)\tau} \right)}{(\mu + \beta) \left( 1 - (1 - r)e^{-(\mu + \beta)\tau} \right)} - \nu - \mu < 0.$$

We thus define a new quantity, the *impulsive reproduction number* 

$$T_0 = \frac{\beta \mu \left( 1 - e^{-(\mu + \beta)\tau} \right)}{(\nu + \mu)(\mu + \beta) \left( 1 - (1 - r)e^{-(\mu + \beta)\tau} \right)},$$

which has the condition that the disease will be controlled if  $T_0 < 1$ .

Solving the equation  $T_0 = 1$ , we can define the maximal period as

$$\hat{\tau} = \frac{1}{\mu+\beta} \ln \frac{(1-r)(\nu+\mu)(\mu+\beta) - \beta\mu}{(\nu+\mu)(\mu+\beta) - \beta\mu}.$$

This is defined only if

$$r < r^* \equiv 1 - \frac{\beta\mu}{(\nu+\mu)(\mu+\beta)}.$$
(1)

Differentiating, we have

$$\frac{\partial T_0}{\partial r} = \frac{\beta \mu \left(1 - e^{-\mu + \beta \tau}\right)}{(\nu + \mu)(\mu + \beta)} \left[ -\left(1 - (1 - r)e^{-(\mu + \beta)\tau}\right) e^{-(\mu + \beta)\tau} \right] < 0.$$

It follows that  $T_0$  is decreasing as r increases, for  $r < r^*$ .

Now let  $r = r^* + \epsilon$  in order to determine what happens beyond  $r^*$ . We have

$$r = \frac{(\nu + \mu)(\mu + \beta) - \beta\mu}{(\nu + \mu)(\mu + \beta)} + \epsilon.$$

Substituting into q and taking a common denominator, we find that the numerator of q is

$$(\nu + \mu)^2 (\mu + \beta)^2 \left[ (1 - \epsilon) e^{-(\mu + \beta)\tau} - 1 \right] < 0.$$

It follows that  $T_0 < 1$  whenever  $r > r^*$ .

In summary, assuming the number of infected vaccinated individuals is negligible, if  $r > r^*$ , where  $r^*$  is defined by (1), then the disease will be controlled, whereas if  $r < r^*$ , then the disease can be controlled, assuming the period between vaccinations satisfies  $\tau < \hat{\tau}$ .

# 5 Numerical simulations

From Weber *et al.* [39], we have  $\beta = 0.03$ ,  $\mu = 0.041$  and  $\nu = 36$ . We add vaccination parameters  $\omega = 0.1$ ,  $\epsilon = 1$ , p = 1,  $\nu_V = 177$  and  $\beta_V = 3000$ . (We also have  $\gamma = 1.8$  and impose  $\gamma_V = 1.2\gamma$ .) This represents a vaccine with complete coverage and perfect efficacy that wanes after ten years, but vaccinated individuals can be infected with a high transmission rate, but recover very quickly.

Figure 3 shows the results of transmission using data from Weber *et al.* [39] and assumed vaccination parameters such that recovery was slightly faster and transmission slightly less likely. The vaccine was given to 50% of the eligible population, but waned after 0.01 years (3.65 days). The data used were  $\mu = 0.041; \omega = 100; \beta = 50; \beta_V = 0.8\beta; \epsilon = 0.9; p = 0.5; \nu = 36; \nu_V = 1.2\nu; \gamma = 1.8; \gamma_V = 1.2\gamma.$ 



Figure 3: Results from the basic model with vaccination. There is an outbreak and the disease oscillates, eventually approaching an equilibrium. A small proportion of individuals are (and remain) vaccinated, with a low-level outbreak among vaccinated individuals.

Figure 4 illustrates the destabilisation of the DFE when extreme vaccination parameters are used. In this case, transmission of the vaccinated strain was extremely high but recovery extremely fast, allowing for infection spikes to occur among a small proportion of vaccinated individuals before the infection stabilises. Data used were  $\mu = 0.041; \omega = 0.1; \beta = 0.03; \beta_V =$  $3000; \epsilon = 1; p = 0, 1; \nu = 36; \nu_V = 177; \gamma = 1.8; \gamma_V = 1.2.\gamma.$ 

Although the transmission rate is unrealistically high, this nevertheless demonstrates that a stable DFE can be destabilised by a vaccine. Note that what we are dealing with here is not a backward bifurcation, but rather a destabilisation of the equilibrium.

Next, following Weber *et al.* [39], we examined the more realistic case



Figure 4: Extreme parameters show that perfect vaccination can induce infection spikes. A. With no vaccine, the result is that the infection clears and the entire population remains susceptible (note that the low-level fluctuations result from numerical limitations in MATLAB) B. With a vaccine given to the entire population, the susceptible population dips slightly as infection takes hold. C. Infection in the vaccinated population initially takes the form of infection spikes before stabilising. Note that vaccination thus destabilises the disease-free equilibrium.

when the transmission rate oscillated. Since the waning rate of the vaccine was not known, we decided to investigate several options for  $\omega$ .

When there is no vaccine, the disease results in a maximum of 7% of the population infected. Data used was  $\mu = 1/70$ ;  $\omega = 1/10$ ;  $b_0 = 60$ ;  $b_1 = 0.16$ ;  $\phi = 0.15$ ;  $\beta_V = 0.5\beta$ ;  $\epsilon = 1$ ; p = 0;  $\nu = 36$ ;  $\nu_V = 1.2\nu$ ;  $\gamma = 1.8$ ;  $\gamma_V = 1.2\gamma$ . See Figure 5.

A vaccine given to the entire population with 50% transmission that did not wane for ten years resulted in about 6% of the population infected. Data used was identical to Figure 5 except that p = 1. See Figure 6. In this case, there is only a slight decrease in the maximum disease burden, despite complete vaccination coverage.

A vaccine given to the entire population with 50% transmission that did not wane for 70 years resulted in a significant reduction in the infected population. Data used was identical to Figure 6 except that  $\omega = 1/70$ . See



Figure 5: Without vaccination, the disease infects up to 7% of the population. A. The total infected population, including vaccinated individuals. B. The final size in each population.



Figure 6: Complete coverage with a vaccine that did not wane for 10 years results in a 1% reduction in the disease compared to not vaccinating. A. The total infected population, including vaccinated individuals. B. The final size in each population.

Figure 7. In this case, there is a significant reduction in the total disease burden, reducing the maximum to less than 2% of the total population.

Note that, even with perfect coverage with a lifelong vaccine (so that  $\epsilon = p = 1$  and  $\mu = \omega = \frac{1}{70}$ ), the DFE still satisfies

$$\bar{S} = \frac{\omega}{\mu + \omega} = \frac{1}{2}$$
$$\bar{V} = \frac{\mu}{\mu + \omega} = \frac{1}{2},$$

so the population without infection would eventually split into equal numbers of vaccinated and unvaccinated susceptible individuals. With infection included and oscillating transmission, explicitly calculating the final size in



Figure 7: Complete coverage with a vaccine that did not wane for 70 years results in a significant reduction in infection. A. The total infected population, including vaccinated individuals. B. The final size in each population.

each compartment is not possible. However, we expect that higher coverage with a lifelong vaccine would tend to a final size with similar numbers; Figure 7 shows that this is indeed the case.

Of course, complete vaccination coverage is not realistic. Consequently, we examined the effect of 50% coverage with a vaccine that did not wane for 70 years. Data used was identical to Figure 7 except that p = 0.5. See Figure 8. In this case, there is still a significant reduction in total disease burden. Note that significantly greater reduction is achieved with 50% coverage and a lifelong vaccine than was achieved with 100% coverage and a vaccine that lasted 10 years (see Figure 6).



Figure 8: 50% coverage with a vaccine that did not wane for 70 years results in a moderate reduction in infection. A. The total infected population, including vaccinated individuals. B. The final size in each population.

It follows that the waning rate of the vaccine is crucial. Even if complete coverage could be achieved, a vaccine with a moderate duration (eg 10 years) results in very little reduction of infection. Conversely, a vaccine that does not wane over a lifetime results in significant reduction in disease burden.

The best-case scenario involves complete coverage with a vaccine that does not wane for 70 years. Figure 9 illustrates the population dynamics when such a vaccine is introduced.



Figure 9: Population dynamics for a lifelong vaccine with complete coverage. Note that the vaccinated infected are too small to appear on the figure.

Figure 10 illustrates the effect of pulse vaccination on the dynamics over a ten-year period. The infection is kept low, with a small outbreak among the vaccinated. Overall, however, there is an exchange with the majority of individuals gradually transforming from susceptible to vaccinated.



Figure 10: Population dynamics for an impulsive vaccine given annually to 50% of the population, with a waning of 10 years. Note that the seasonal oscillations and the impulses combine to produce a double period.

### 6 Discussion

The introduction of a vaccine is always desirable, but new vaccines pose the risk of unintended consequences. We have highlighted some of the potential issues that may arise with vaccination against RSV. In particular, we determine conditions under which a destabilisation of the disease-free equilibrium is possible. This is not in the form of a backward bifurcation, as is sometimes seen, but rather occurs when the vaccine causes sufficiently fast recovery and transmission is extremely high. An infection-free population that is effectively vaccinated against RSV can nevertheless produce vaccination-innduced spikes of infection. Although such a case is unlikely to occur with the unrealistic parameters we chose, we have shown proof-ofconcept that it is possible and determined conditions on the recovery rate due to vaccinaton that allow for the possibility.

Using impulsive differential equations, we were able to formulate conditions on the period and the strength of vaccination to allow for disease control (though not eradication). If the vaccine reduces transmissibility and is applied frequently, then vaccinated infected individuals can be reduced to low numbers. We relaxed the assumption of constant transmission. We demonstrated that the waning of the vaccine has a greater effect on the outcome that coverage. Hence it is imperative that a good vaccine be developed before being released for general use.

We also defined a new quantity, the impulsive reproduction number  $T_0$ . This is a sufficient (but not necessary) condition, based on an overestimate of the infected population, that ensures eradication if  $T_0 < 1$ . If  $T_0 < 1$ , then the infected population is contracting within each impulsive cycle. Since the infected population is then reduced at each impulse point, the result is the eventual eradication of the infection. Note that we assumed constant transmission for this derivation; however, numerical simulations were performed using seasonal oscillations. The result was a double period: one from the impulsive periodic orbit and the other from the seasonal oscillations.

Our model has some limitations, which should be acknowledged. We assumed that time to administer the vaccine was significantly shorter than the time between vaccine administrations in order to justify the impulsive approximation. Such assumptions are reasonable in many cases [30], although can produce confounding effects in some situations [11]. The extreme parameters that we used to illustrate the vaccination spikes operated under the assumption that the transmission rate for infected vaccinatied individuals was significantly higher than the transmission rate without vaccination. Since we extended the model of Weber *et al.* [39], our model inherited many of the assumptions from that model, such as mass-action transmission, a constant birth rate and that the birth and death rates were matched, resulting in a constant population.

A vaccine that targets RSV infection has the potential to significantly reduce the overall prevalence of the disease, but it has to be sufficiently long-lasting. Coverage and effectiveness of the vaccine is important, but the critical parameter that our modelling identified is the waning rate of the vaccine. We thus recommend that candidate vaccines be tested for sufficient duration before being released to the public. If a durable vaccine can be developed, then we stand a chance of controlling this disease, assuming sufficiently widespread coverage.

## References

- Acedo, L., J. Díez-Domingo, J.-A. Moraño, and R.-J. Villanueva (2010). Mathematical modelling of respiratory syncytial virus (RSV): vaccination strategies and budget applications. *Epidemiology and Infection* 138(6), 853–60.
- [2] Acedo, L., J.-A. Moraño, and J. Díez-Domingo (2010). Cost analysis of a vaccination strategy for respiratory syncytial virus (RSV) in a network model. *Mathematical and Computer Modelling* 52(7-8), 1016–1022.
- [3] Arenas, A. J., G. González-Parra, and L. Jódar (2010). Randomness in a mathematical model for the transmission of respiratory syncytial virus (). Mathematics and Computers in Simulation 80(5), 971–981.
- [4] Avendano, L. F., M. Ange, and C. Larran (2003). Surveillance for Respiratory Syncytial Virus in Infants Hospitalized for Acute Lower Respiratory Infection in Chile (1989 to 2000). *Journal of Clinical Microbiol*ogy 41(10), 4879–4882.
- [5] Bainov, D. and P. Simeonov (1989). Systems with Impulsive Effect. Ellis Horwood Ltd.
- [6] Bainov, D. and P. Simeonov (1993). *Impulsive differential equations:* periodic solutions and applications. Longman Scientific and Technical.
- [7] Bainov, D. and P. Simeonov (1995). Impulsive Differential Equations: Asymptotic Properties of the Solutions. World Scientific.

- [8] Brandenburg, A. H., J. Groen, H. A. van Steensel-Moll, E. C. Claas, P. H. Rothbarth, H. J. Neijens, and A. D. Osterhaus (1997). Respiratory syncytial virus specific serum antibodies in infants under six months of age: limited serological response upon infection. *Journal of Medical Virology* 52(1), 97–104.
- [9] Cane, P. A. (2001). Molecular epidemiology of respiratory syncytial virus. *Reviews in Medical Virology* 11(2), 103–116.
- [10] Capistran, M., M. Moreles, and B. Lara (2009). Parameter Estimation of Some Epidemic Models. The case of recurrent epidemics caused by respiratory syncytial virus. *Bulletin of Mathematical Biology* 71, 1890– 1901.
- [11] Church, K. and R. J. Smith? (2014). Analysis of piecewise-continuous extensions of periodic linear impulsive differential equations with fixed, strictly inhomogeneous impulses. Dynamics of Continuous, Discrete and Impulsive Systems, Series B: Applications & Algorithms 21, 101–119.
- [12] Domachowske, J. B. and H. F. Rosenberg (1999). Respiratory Syncytial Virus Infection: Immune Response, Immunopathogenesis, and Treatment. *Clinical Microbiology Reviews* 12(2), 298–309.
- [13] Fields, B. S., B. L. House, J. Klena, L. W. Waboci, T. Whistler, and E. C. Farnon (2013). Role of global disease detection laboratories in investigations of acute respiratory illness. *The Journal of Infectious Diseases 208 Suppl*(Suppl 3), S173–176.
- [14] Hall, C. B. (1981). Respiratory syncytial virus. In R. D. Feigin and J. D. Cherry (Eds.), *Textbook of Paediatric Infectious Diseases, 1st edn* (*Volume II*), Chapter 28 - Viral, pp. 1247–1267. Philadelphia; London: W. B. Saunders Company.
- [15] Hall, C. B. (2001). Respiratory syncytial virus and parainfluenza virus. New England Journal of Medicine 344 (25), 1917–1928.
- [16] Han, L. L., J. P. Alexander, and L. J. Anderson (1999). Respiratory syncytial virus pneumonia among the elderly: an assessment of disease burden. *The Journal of Infectious Diseases* 179(1), 25–30.
- [17] Hardelid, P., R. Pebody, and N. Andrews (2013). Mortality caused by influenza and respiratory syncytial virus by age group in England and Wales 1999-2010. *Influenza and Other Respiratory Viruses* 7(1), 35–45.

- [18] Haynes, L. M. (2013). Progress and Challenges in RSV Prophylaxis and Vaccine Development. *The Journal of Infectious Diseases 208* Suppl(Suppl 3), S177–183.
- [19] Henderson, F. W., A. M. Collier, W. A. Clyde Jr, and F. W. Denny (1979). Respiratory-Syncytial-Virus Infections, Reinfection and Immunity: A Prospective, Longitudinal Study in Young Children. *The New England Journal of Medicine* 300(10), 530–534.
- [20] La Via, W., M. Marks, and H. Stutman (1992). Respiratory syncytial virus puzzle: clinical features, pathophysiology, treatment, and prevention. *The Journal of Pediatrics 121*(4), 503–510.
- [21] Lakshmikantham, V., D. Bainov, and P. Simeonov (1989). Theory of Impulsive Differential Equations. World Scientific.
- [22] Leecaster, M., P. Gesteland, T. Greene, N. Walton, A. Gundlapalli, R. Rolfs, C. Byington, and M. Samore (2011). Modeling the variations in pediatric respiratory syncytial virus seasonal epidemics. *BMC Infectious Diseases* 11(1), 105.
- [23] McNamara, P. S. and R. L. Smyth (2002). The pathogenesis of respiratory syncytial virus disease in childhood. *British Medical Bulletin* 61, 13–28.
- [24] Meng, J., C. C. Stobart, A. L. Hotard, and M. L. Moore (2014). An overview of respiratory syncytial virus. *PLoS pathogens* 10(4), e1004016.
- [25] Mlinaric-Galinovic, G., R. C. Welliver, T. Vilibic-Cavlek, S. Ljubin-Sternak, V. Drazenovic, I. Galinovic, and V. Tomic (2008). The biennial cycle of respiratory syncytial virus outbreaks in Croatia. *Virology Journal* 5(18).
- [26] Moore, H. C., P. Jacoby, A. B. Hogan, C. C. Blyth, and G. N. Mercer (2014). Modelling the Seasonal Epidemics of Respiratory Syncytial Virus in Young Children. *PLoS ONE* 9(6), e100422.
- [27] Nair, H., D. J. Nokes, B. D. Gessner, M. Dherani, S. A. Madhi, R. J. Singleton, K. L. O'Brien, A. Roca, P. F. Wright, N. Bruce, A. Chandran, E. Theodoratou, A. Sutanto, E. R. Sedyaningsih, M. Ngama, P. K. Munywoki, C. Kartasasmita, E. A. F. Simões, I. Rudan, M. W. Weber, and H. Campbell (2010). Global burden of acute lower respiratory infections due to respiratory syncytial virus in young children: a systematic review and meta-analysis. *Lancet* 375(9725), 1545–55.

- [28] Panozzo, C. A., A. L. Fowlkes, and L. J. Anderson (2007). Variation in timing of respiratory syncytial virus outbreaks: lessons from national surveillance. *The Pediatric Infectious Disease Journal* 26(11 Suppl), S41– 45.
- [29] Paynter, S., L. Yakob, E. A. F. Simões, M. G. Lucero, V. Tallo, H. Nohynek, R. S. Ware, P. Weinstein, G. Williams, and P. D. Sly (2014). Using mathematical transmission modelling to investigate drivers of respiratory syncytial virus seasonality in children in the Philippines. *PLoS ONE* 9(2), e90094.
- [30] Smith?, R. J. and E. J. Schwartz (2008). Predicting the potential impact of a cytotoxic t-lymphocyte hiv vaccine: How often should you vaccinate and how strong should the vaccine be? *Mathematical Biosciences 212*, 180–187.
- [31] Sorce, L. R. (2009). Respiratory syncytial virus: from primary care to critical care. *Journal of Pediatric Health Care* 23(2), 101–108.
- [32] Stensballe, L. G., J. K. Devasundaram, and E. A. F. Simoes (2003). Respiratory syncytial virus epidemics: the ups and downs of a seasonal virus. *Pediatric Infectious Disease Journal* 22(2), S21–S32.
- [33] Sullender, W. M. (2000). Respiratory syncytial virus genetic and antigenic diversity. *Clinical Microbiology Reviews* 13(1), 1–15.
- [34] Terletskaia-Ladwig, E., G. Enders, G. Schalasta, and M. Enders (2005). Defining the timing of respiratory syncytial virus (RSV) outbreaks: an epidemiological study. *BMC Infectious Diseases* 5(20).
- [35] The IMpact-RSV Study Group (1998). Palivizumab, a Humanized Respiratory Syncytial Virus Monoclonal Antibody, Reduces Hospitalization From Respiratory Syncytial Virus Infection in High-Risk Infants. *Pediatrics* 102(3), 531–537.
- [36] Tregoning, J. S. and J. Schwarze (2010). Respiratory viral infections in infants: causes, clinical symptoms, virology, and immunology. *Clinical Microbiology Reviews* 23(1), 74–98.
- [37] van Asten, L., C. van den Wijngaard, W. van Pelt, J. van de Kassteele, A. Meijer, W. van der Hoek, M. Kretzschmar, and M. Koopmans (2012).

Mortality attributable to 9 common infections: significant effect of influenza A, respiratory syncytial virus, influenza B, norovirus, and parainfluenza in elderly persons. *The Journal of Infectious Diseases 206*(5), 628–639.

- [38] Wang, E. E. and B. J. Law (1998). Respiratory syncytial virus infection in pediatric patients. Seminars in Pediatric Infectious Diseases 9(2), 146– 153.
- [39] Weber, A., M. Weber, and P. Milligan (2001). Modeling epidemics caused by respiratory syncytial virus (RSV). *Mathematical Bio-sciences* 172(2), 95–113.
- [40] Weber, M. W., E. K. Mulholland, and B. M. Greenwood (1998). Respiratory syncytial virus infection in tropical and developing countries. *Tropical Medicine and International Health* 3(4), 268–280.
- [41] White, L. J., J. N. Mandl, M. G. M. Gomes, A. T. Bodley-Tickell, P. A. Cane, P. Perez-Brena, J. C. Aguilar, M. M. Siqueira, S. A. Portes, S. M. Straliotto, M. Waris, D. J. Nokes, and G. F. Medley (2007). Understanding the transmission dynamics of respiratory syncytial virus using multiple time series and nested models. *Mathematical Biosciences 209*(1), 222–239.
- [42] Yorita, K. L., R. C. Holman, C. A. Steiner, P. V. Effler, J. Miyamura, S. Forbes, L. J. Anderson, and V. Balaraman (2007). Severe bronchiolitis and respiratory syncytial virus among young children in Hawaii. *The Pediatric Infectious Disease Journal* 26(12), 1081–1088.